Water quality dynamics of ephemeral wetlands in the Piedmont ecoregion, South Carolina, USA

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ABSTRACT

Small ephemeral wetlands are commonly found in the Piedmont ecoregion of the southeastern USA. Ephemeral wetlands have important ecological functions but information about their water quality over its flooding periods is relatively limited. In this study, the water chemistry and physical parameters of three ephemeral wetlands and their nearby water bodies, including first order and second order streams and groundwater in the Piedmont ecoregion of South Carolina, were closely monitored during their flooding periods from January to June 2012. Nutrient and water quality analyses demonstrated the chemistries of wetlands, stream, and groundwater were different from each other in spite of their proximity. Greater concentrations of dissolved organic carbon (DOC) and dissolved organic nitrogen with a major portion in humic-acid-like and fulvic-acid-like fractions were generally found in wetland waters. In contrast, significantly lower DOC concentrations with a greater portion of inorganic nitrogen were observed in stream and groundwater. Electrical conductivity at 25°C (EC\textsubscript{25}) and temperature measurements showed a greater fluctuation in wetlands, indicating their poor buffering capacity against environmental changes. Results of this field study suggested that these small ephemeral wetlands in the Piedmont Ecoregion have relatively unique biogeochemistry in comparing their adjacent water bodies.

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1. Introduction

Ephemeral wetlands (i.e., isolated, seasonal, temporary, dry end, or headwater wetlands; vernal pools) provide numerous watershed-level functions including critical wildlife habitat, floodwater storage, groundwater recharge, and water filtration. Global declines in ephemeral wetlands have been linked to amphibian declines, loss of habitat for reptiles and invertebrates, and alterations to hydrological regimes (Gibbons et al., 2000; Jenkins et al., 2003; Zedler and Kercher 2005). Smaller wetlands are less able to recover functions after human disturbance (Moreno-Mateos et al., 2012), yet because of their dispersion in the landscape provide value disproportionate to their size (Gibbs, 1993; Leibowitz 2003). Despite the importance of these wetlands, they are poorly protected in much of the United States and have, at best, ambiguous levels of regulatory oversight under the CWA (Zedler, 2003). U.S. Supreme Court decisions (e.g., Rapanos et ux et al. v. United States, 547 U.S. 715, 2006) suggested that water bodies other than traditional navigable waters (TNWs) and the adjacent wetlands and relative permanent tributaries of TNWs and the abutting wetlands could be jurisdictional waters if a significant nexus based on hydrological or ecological connectivity existed with a TNW (Grumbles et al., 2008). A recent U.S. rule expands the definition of waters of the United States and establishes a pressing need for better scientific information to define connectivity among water bodies (Federal Register, 2014). The Piedmont ecoregion of the southeastern United States is a useful instance of loss of wetlands to historical agriculture that had intensive impacts on geomorphology of aquatic systems; such have been compounded by recent urbanization and land-use changes (Campbell et al., 2008; Napton et al., 2010). The extent of wetland loss is largely unknown and, until recently, few small, ephemeral, ‘isolated’ wetlands in the Piedmont ecoregion had been mapped, let alone studied (Pitt et al., 2012). Population growth, land use trends, and the resultant pressure on wetlands and other aquatic resources has created a need to better understand small, ephemeral, ‘isolated’ wetlands.

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Different methods and characteristics have been used to examine the connectivity of wetlands and their nearby water bodies, including physical measurements i.e., water level and temperature, (Cabezas et al., 2011; Glinka-Lewczuk, 2009), chemical parameters such as nitrogen and phosphorus (Wolf et al., 2013; Glinka-Lewczuk, 2009), biological indicators such as macrozoobenthos patterns (Obolewski, 2011), as well as mathematical modeling (Golden et al., 2014). These methods, generally used in floodplain or riparian wetlands, may not be directly applicable to more geographically isolated, ephemeral wetlands in the Piedmont ecoregion because of the wetlands’ unique physical characteristics including small size, shallow depth, and relatively short hydroporid.

In order to understand the connectivity and ecological function of Piedmont ephemeral wetlands, we first need to understand their water quality dynamics through the entire hydroporid period. In this study, we examined the water chemistry and physical parameters of three ephemeral wetlands and the nearby water bodies between 26 January and 5 June 2012, a time period that encompassed the typical flooded period of ephemeral wetlands in Piedmont ecoregion of the southeastern United States. In addition to general water quality and nutrient analyses, we introduce the use of optical properties of dissolved organic matter (DOM) to examine ephemeral wetland biogeochemistry compared to nearby water bodies. DOM produced from different sources has a unique optical signature in ultraviolet and visible light absorption and fluorescence spectra and has been used in source-water tracking (Osburn et al., 2012; Chow et al., 2008). We hypothesized that small, ephemeral, ‘isolated’ wetlands have unique biogeochemical characteristics in comparing with nearby permanent water bodies which had different hydrology. We designed our study to contribute to the growing understanding of how to evaluate chemical relationships among water bodies in the context of “Significant Nexus.”

2. Materials and methods

2.1. Study sites

We selected two index landscapes within the Piedmont ecoregion of South Carolina, USA (Fig. 1a and b). The first index landscape contained a first order stream, two ephemeral wetlands with varying degrees of geographic isolation, and a cypress swamp with surficial hydrological connections with the stream and a lake. Site A within the first index landscape (Fig. 1c) had the smallest ephemeral wetland with a maximum surface area of 6.75 m² and it was approximately 10.7 m from the perennial stream. The ephemeral wetland in site B (Fig. 1c) of the first index landscape had a maximum surface area of 37.74 m² and was adjacent to a formerly ephemeral wetland that became permanent and surficially hydrologically connected with the stream due to beaver damming activity. Site B also contained the cypress swamp with surficial hydrological connections to the stream and lake (Fig. 1c). The second index landscape contained site C (Fig. 1d) which included a second order stream, an ephemeral floodplain wetland, and a marsh with surficial hydrological connections with a lake. The wetland in site C had a maximum surface area of 244.40 m², excluding the areas that became flooded for a short duration (<1 week) during flooding events of the adjacent stream.

2.2. Piezometer construction and water level measurement

Piezometers were installed on 8–10 January 2012 in order to sample groundwater. The locations of the piezometers were selected based on their relative positions between the ephemeral wetlands and nearby streams (Figs. SI-1–SI-3, as available in Supplementary Information [SI]). For constructing a piezometer, a 15 cm-diameter x 50 cm-depth borehole was drilled. A 1 m-long x 5 cm-diameter PVC pipe was placed in the borehole. The bottom of the PVC pipe was capped with a slotted screen 30 cm from the bottom. Gravel was placed around the slotted interval and bentonite was added on top to prevent water infiltrating from the soil surface. Levels of groundwater (denoted as G) were manually measured using a portable water level meter. Depths of wetlands (denoted as W) and streams (denoted as S) at selected locations were determined manually using meter sticks. Rain gauges (denoted as R) were placed in each site for precipitation determination. There are a total 9 sampling points in Site A (i.e., 5 groundwater denoted as A-G1 to A-G5; 2 wetland waters denoted as A-W1 and A-W2; 2 stream waters denoted as A-S1 and A-S2), 13 sampling points in Site B (i.e., 5 groundwater denoted as B-G1 to B-G5; 5 wetland waters denoted as B-W1 to B-W5; 3 stream waters denoted as B-S1 to B-S3), 16 sampling points in site C (i.e., 10 groundwater denoted as C-G1 to C-G10; 3 wetland waters denoted as C-W1 to C-W3; 3 stream waters denoted as C-S1 to C-S3). Measurements were conducted at least three times per week during the study period. Rates of change in water level or water depth (Δh/Δt) at each point were calculated by dividing the difference between two measurements by the time interval.

2.3. Water quality determination

General water quality of stream and wetland waters was determined using YSI 556 Multiprobe System equipped with dissolved oxygen (DO), pH, electrical conductivity corrected to 25 °C (EC25), turbidity, oxidation-reduction potential (ORP), and temperature sensors (YSI, Inc., Yellow Springs, OH, USA). The probe was gently placed 5–10 cm below the water surface to minimize any disturbance of wetland sediments. Measurements were generally taken between 0900 and 1500 h and these field surveys were conducted at least three times per week. Grab samples were collected once a month for nutrient analysis and dissolved organic matter (DOM) characterization (Section 2.4). Surface water, approximately 5 cm below the water’s surface, was collected in 125 mL pre-acid washed polyethylene bottles. Soil pore water was manually pumped from the pre-installed piezometers (Section 2.2). At least one liter of water was pumped and discarded from the piezometer before collecting into a 125 mL bottle. Each sample type (i.e., wetland, stream, and groundwater) had at least two sampling spots and 3 sampling months, and the sample size was always ≥6. All samples were immediately stored in an ice cooler and transported to the laboratory. Waters were then filtered through 0.45 μm membrane filters (Millipore Express PLUS Membrane, polyethersulfone, hydrophilic, 47 mm). Filtrates were then stored at 4°C until further analysis. All measurements and samples were collected between 26 January and 15 June 2012. Water quality measurements ended when the ephemeral wetlands were dry. Only one wetland in site B was fully examined for six months. The wetlands in sites A and C (Fig. 1c and d) were completely dry in April.

2.4. Chemical analyses

Each filtered sample was analyzed for dissolved organic carbon (DOC) and total dissolved nitrogen (TDN) using a Shimadzu TOC/TN analyzer. Inorganic N, including NH₄-N and (NO₂ + NO₃)-N were determined using a Systex® Easychem™ discrete analyzer (EPA Methods 350.1 and 325.2, respectively; Eaton and Franson, 2005). DOC was further characterized by Shimadzu UV-1800 visible and ultraviolet spectrophotometer scanning from 200 to 700 nm. Specific ultraviolet absorbance (SUVA) was calculated by normalizing ultraviolet absorbance at 254 nm to DOC concentration, recorded as L mg C−1 m−1. SUVA has been widely used as a surrogate for aro-
mastic carbon content of DOC (Chow et al., 2008). Spectral slope ratio ($S_R$), which is correlated with molecular weight (MW) of DOC and to photochemically induced shifts in MW, was calculated as the ratio of two spectral slopes between 275 and 295 nm and 350–400 nm (Helms et al., 2008; Loiselle et al., 2009). In addition, the $E_2/E_3$ ratio, absorbance at 254 nm ($E_2$) divided by absorbance at 365 nm ($E_3$), was calculated. The $E_2/E_3$ ratio, which is positively correlated with the quantum yields of singlet oxygen ($^{1}\text{O}_2$) but negatively correlated with hydrogen peroxide ($\text{H}_2\text{O}_2$) (Dalrymple et al., 2010), is an important index showing the photo-degradability and photo-reactivity of DOC. Selected samples were further analyzed using fluorescence emission-excitation matrix (EEM). The components of DOC were quantified using the fluorescence regional integration (FRI) model that separates into aromatic protein I, aromatic protein II, fulvic-acid-like, microbial-byproduct-like, and humic-acid-like fractions (Zhou et al., 2013). Samples were diluted with deionized water to the absorbance at 254 nm to 0.3 or lower, and were measured using a Shimadzu RF-5301 spectrofluorophotometer. Ultraviolet-visible (UV/VIS) and fluorescence spectroscopy have been used to track the origins and the sources of DOM in water (Chow et al., 2008; Osburn et al., 2012).

2.5. Statistical analyses

Water quality parameters among different water bodies were compared using a paired-samples t-test. In addition to the average and 95% confidence limits of measured values, daily differences were calculated in order to eliminate the seasonal effects. Parameters of two water bodies were considered the same if their differences were not significantly different from zero. For those parameters that did not conform to a normal distribution (Kolmogorov-Smirnov test) and homogeneity of variance (Levene test), a Wilcoxon signed rank test was used. To detect differences in water quality parameters at different times, ANOVA and a post-hoc Tukey's test were used unless the non-parametric comparison was specified. A Kruskal–Wallis test and a non-parametric Tukey-type multiple comparison tests were used when parameters did not conform to a normal distribution and homogeneity of variance. Statistical methods were implemented using SPSS version 19.0 (IBM Corporation, Armonk, NY, USA) and the level of significance was $\alpha = 0.05$.

3. Results

3.1. Water level and hydroperiod

The wetland in site A had the smallest surface area (6.75 m²) but the greatest water depth (40.3 cm); it was inundated for approximately 4 months and was completely dry by late April. The wetland in site B had a surface area of 37.74 m² with a water depth of 34.8 cm. The wetland was inundated for over 6 months and was still flooded at the end of the study period (15 June 2013). The wetland in site C had the largest surface area of the three studied wetlands (2444 m²) but the shallowest depth (27.1 cm). Most of the area in this wetland was dry in early February but some areas were still flooded until late April.

The water levels of these wetland systems were generally sensitive to precipitation. Rapid increases in water levels were observed during rainstorms. For example, sudden increases of water levels in groundwater and wetland were observed in site A on 3 March, 17 April, and 15 May, when intensive rainstorms (>0.5 mm per day) occurred (Fig. 2). Notably, both groundwater and wetland levels

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**Fig. 1.** (a) The Piedmont ecoregion (depicted by gray) of South Carolina, USA. Star indicates the location of the study area. (b) Map of the two index landscapes used to evaluate hydrological and biogeochemical connectivity. (c) Index landscape 1 includes site A and B. (d) Index landscape 2 includes site C.
showed similar magnitude of changes. Following rain events, they each rapidly dropped back to their background levels. Although there were slight increases in water depth of nearby streams, the levels varied <10% during the 6-month study period.

Correlation coefficient matrices among the rates of change in water level \((\Delta h/\Delta t)\) calculated from different sampling points were compared and summarized in Tables SI-1–SI-3. In site A, \(\Delta h/\Delta t\) of wetland, groundwater, and stream showed strong correlations. Correlation coefficients of 0.8 or greater were recorded between stream (A-S1 and A-S2, denoted as site A stream points 1 and 2, respectively) and groundwater (A-G2, A-G3, A-G4, and A-G5, denoted as site A groundwater points 2, 3, 4, and 5, respectively). Correlation coefficients of 0.6 or greater between wetland (A-W1 and A-W2, denoted as site A wetland points 1 and 2, respectively) and stream (A-S1 and A-S2), and between wetland (A-W2) and groundwater (A-G5) were reported (Table SI-1). In contrast to site A, the correlations among different points in site B were relatively weaker. Although few points such as B-S2 versus B-G4 and B-W4 versus B-W2 had correlation coefficients of 0.7 or greater, many correlation coefficients were below 0.3 (Table SI-2). Results of site C were similar to that of site A. Strong correlations among wetland, groundwater, and stream were observed. Except few sampling points such as C-W1 and C-G8, many of the correlation coefficients were above 0.6.

3.2. General water quality

3.2.1. Site A

Water temperatures of the stream and the ephemeral wetland in site A were closely matched during the 3-month study period but the range of fluctuation in the wetland was greater than that in the stream (Fig. 3a). The stream fluctuated between 6.2 and 13.6 °C \((\Delta\text{Temp} = 7.4 °C)\) whereas the wetland fluctuated between 3.2 and 15.3 °C \((\Delta\text{Temp} = 12.1 °C)\). In contrast to the temperature, the water chemistry of the stream and wetland in site A showed obvious differences. The electrical conductivity \((\text{EC}_{25})\) of the stream \((16.45 \pm 2.59 \mu\text{S cm}^{-1}, \text{average } \pm \text{standard deviation})\) was significantly lower \((p < 0.05; n = 20; t\text{-test})\) than that of the wetland \((34.20 \pm 4.11 \mu\text{S cm}^{-1}; \text{Fig. 3b})\). A greater pH value \((p < 0.05)\) was generally observed in the stream, with an average of 6.53 ± 0.24, comparing to the pH of the wetland, which had an average of 6.12 ± 0.11 (Fig. 3c). Dissolved oxygen \((\text{DO})\) in the stream was also consistently higher with nearly 100% saturation, but DO in the wetland was typically below 50% (Fig. 3d).

Similarly to water temperature, water quality parameters of the stream were relatively stable compared to that of the wetland. \(\text{EC}_{25}\) and DO of the stream fluctuated in a relatively narrow range \((\Delta\text{EC}_{25} = 9.5 \mu\text{S cm}^{-1} \text{ and } \Delta\text{DO} = 12.85%), \text{compared to those in the wetlands } (\Delta\text{EC}_{25} = 13.5 \mu\text{S cm}^{-1} \text{ and } \Delta\text{DO} = 43.6%)\). To eliminate the effects of temporal variation, the differences of daily measurements between stream \((X_1)\) and wetland \((X_2)\) at each sampling event were calculated, as summarized in Table SI-4. Except the temperature, all measures (pH, DO, and \(\text{EC}_{25}\)) were significantly different between wetland and stream waters. For the nutrient analysis, the ephemeral wetland showed a greater DOC concentration \((p < 0.05)\) with an average of 4.27 ± 0.29 mg L\(^{-1}\) than that in groundwater and stream but no difference in nitrogen was observed (Table 1). A significantly higher C/N ratio \((13.38 \pm 3.04 \text{ mol mol}^{-1})\) was found in wetland compared to stream \((7.98 \pm 0.80 \text{ mol mol}^{-1})\) or groundwater \((8.62 \pm 1.25 \text{ mol mol}^{-1})\).

3.2.2. Site B

Water temperature, \(\text{EC}_{25}\), pH, and DO of the stream and the ephemeral wetland in site B generally behaved the same as in site A (Figure SI-4). Greater temperature and \(\text{EC}_{25}\), but lower pH and DO were found in the wetland (Table SI-4). In contrast to the wetlands in sites A and C, DOC concentration in the ephemeral wetland in site B was similar to those in the stream and groundwater, with a mean of 1.74 ± 0.11 mg L\(^{-1}\). The C/N ratio in the wetland was less than 10 mol mol\(^{-1}\) and was not significantly different \((p > 0.05)\) from that in the stream. Notably, the wetland in site B was located in an open area with fewer trees in the surrounding landscape. In addition, water quality of a nearby connected wetland (Figure SI-2) was compared to that of the ephemeral wetland, and the results are summarized in Figure SI-5. The water quality between two wetlands had very similar patterns and values, although the ephemeral wetland generally had lower \(\text{EC}_{25}\), pH, and DO. TN, NO\(_3\)-N, NH\(_4\)-N, and DOC concentrations of the connected wetland were similar to the ephemeral wetland and their differences were insignificant \((p > 0.05; \text{Tables 1 and 2})\). Although the connected wetland had obvious surface water connection with the nearby water bodies, obvious water movement was not observed. Its chemistry was quite similar to that of the ephemeral wetland.

3.2.3. Site C

In spite of the differences in surface area and water depth, the general water quality of the ephemeral wetland in site C was similar to those in sites A and B. The temperature of the ephemeral wetland was significantly higher \((p < 0.05)\) than that of the stream water, with a mean of 2.24 °C difference (Table SI-4). The pH of the wetland \((6.72 \pm 0.23)\) was generally lower than that of the stream water \((7.42 \pm 0.36)\). The \(\text{EC}_{25}\) of the wetland was greater with an average of 66 ± 12.5 μS cm\(^{-1}\). The stream \(\text{EC}_{25}\) was only 37 ± 4 μS cm\(^{-1}\). However, the difference in \(\text{EC}_{25}\) was not statistically significant \((p > 0.05)\) due to its greater fluctuation (Figure SI-6). DO of the stream water was very stable with nearly 100% saturation whereas the wetland’s DO was generally below 50%. The wetland in site C had the highest DOC concentration among all water bodies in this study, with a mean of 6.54 ± 2.17 mg L\(^{-1}\). Notably, groundwater in site C had the highest total nitrogen concentration, dominated with NO\(_3\)-N \((0.71 \pm 0.22 \text{ mg L}^{-1})\) but also a significant portion of NH\(_4\)-N \((0.59 \pm 0.01 \text{ mg L}^{-1})\). Stream water in this site, which was a 2nd order stream, also had a substantial level of NO\(_3\)-N \((0.32 \pm 0.01 \text{ mg L}^{-1})\). In contrast to water in the ephemeral wetland, dissolved organic N, as calculated by subtracting TN from NO\(_3\)-N and NH\(_4\)-N, was the major nitrogen species.

3.3. Optical properties of DOM

The ephemeral wetland in sites A and C had significantly greater SUVA values \((p < 0.05)\) than that of stream and groundwater. In site B, no differences were observed in SUVA among three water bodies.
Fig. 3. General water quality of ephemeral wetland and adjacent stream at site A. Water quality was determined in the wet period of the wetland between 1/26/2012 and 4/22/2012. Values and error bars are the average and the range of two measurements.

Table 1
Comparison of monthly nutrient concentrations (mean ± standard deviation, n ≥ 6) among different water bodies in sites A, B, and C. Means with different superscript letter (a, b, and c) in the same row indicate the difference is statistically significant (p < 0.05). All concentrations units are in mg L⁻¹, except C/N ratio, which are expressed in mol mol⁻¹. NP = using non-parametric comparison as the normality tests did not pass.

<table>
<thead>
<tr>
<th>Site</th>
<th>Parameters</th>
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<th>Groundwater</th>
<th>Ephemeral Wetland</th>
<th>Connected Wetland</th>
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<tr>
<td>A</td>
<td>DOC&lt;sup&gt;NP&lt;/sup&gt;</td>
<td>1.73 ± 0.22&lt;sup&gt;b&lt;/sup&gt;</td>
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<td>B</td>
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<td>C/N&lt;sup&gt;NP&lt;/sup&gt;</td>
<td>5.98 ± 2.63&lt;sup&gt;a&lt;/sup&gt;</td>
<td>4.60 ± 2.93&lt;sup&gt;*&lt;/sup&gt;</td>
<td>11.11 ± 1.32&lt;sup&gt;b&lt;/sup&gt;</td>
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</table>
The E2/E3 ratio and spectral slope (S<sub>e</sub>), which are considered surrogate of molecular weight or size of DOM (Helms et al., 2008; Loiselle et al., 2009), showed no statistical differences (p > 0.05) among stream, groundwater, and wetlands in all three sites (Table 2). Noticeably, E2/E3 and S<sub>e</sub> could not be calculated when DOC concentration was too low and absorbance greater than 300 nm could not be detected, such as the groundwater in sites A and B. However, lower SUVA but higher E2/E3 values in waters collected in January were generally observed when comparing to those collected in March.

The fluorescence EEMs of stream, groundwater, and wetland in site A are shown in Fig. 4. All waters showed a strong peak in the areas of excitation (Ex) < 240 nm and 400 nm < emission (Em) < 500 nm, representing fulvic-acid-like compounds. Both stream and wetland waters showed a small peak in Ex = 280 nm and Em = 250 nm, indicating humic-acid-like compounds. This humic-acid-like peak was not obvious in groundwater. In contrast, stream and groundwater showed signals in the regions of Ex < 240 nm and 300 nm < Em < 400 nm, which are considered aromatic protein-like regions. This protein-like peak was very weak in wetlands. A fluorescence regional integration (FRI) using Simpson’s rule (Zhou et al., 2013) was applied to quantify the relative abundance of these fractions, and the results of the monthly samples in site A were summarized in Fig. 5. Fulvic acid-like compounds were the dominated fraction in stream and wetland waters, representing 56 and 42%, respectively. Only 28% of fulvic-acid-like compound was found in groundwater. Groundwater was dominated by aromatic protein compounds, which represent 49% of total DOM. Wetland water had the highest humic-like compounds, with 17%, followed by the stream (12%) and groundwater (10%). The distributions of these 5 operationally defined fractions only slightly changed in the 3-month study period. The increase of aromatic protein region but the decrease in fulvic- and humic-acid-like fractions were observed in wetland and stream waters from January to March. The fraction distribution in groundwater was relatively stable in the 3 months.

4. Discussion

4.1. Physical settings of ephemeral wetlands

The size of a wetland (i.e., surface area and water depth) plays a significant role in determining its water and habitat qualities (Kaplan et al., 2011; Cosentino et al., 2010). The buffer capacity against environmental changes is proportional to the volume of water within an isolated wetland (Brinson, 1993). In our study, a greater fluctuation in water quality in these small wetlands, such as temperature and EC<sub>5</sub>, was generally observed compared to their adjacent water bodies. The differences in water tempera-

Table 2
Comparison of monthly optical properties of DOM (mean ± standard deviation, n = 3) among different water bodies in sites A, B, and C. Means with different superscript letter (a, b, and c) in the same row indicate the difference is statistically significant (p < 0.05). NP = using non-parametric comparison. N.A. = Not applicable.

<table>
<thead>
<tr>
<th>site A.</th>
<th>Parameters</th>
<th>Stream</th>
<th>Groundwater</th>
<th>Ephemeral Wetland</th>
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</thead>
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<tr>
<td>SUVA&lt;sub&gt;0.69&lt;/sub&gt;</td>
<td>1.64 ± 0.16&lt;sup&gt;b&lt;/sup&gt;</td>
<td>0.32 ± 0.01&lt;sup&gt;a&lt;/sup&gt;</td>
<td>3.32 ± 0.26&lt;sup&gt;a&lt;/sup&gt;</td>
<td></td>
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<tr>
<td>E2/E3</td>
<td>3.65 ± 0.18&lt;sup&gt;b&lt;/sup&gt;</td>
<td>N.A.</td>
<td>4.10 ± 0.42&lt;sup&gt;b&lt;/sup&gt;</td>
<td></td>
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<tr>
<td>S&lt;sub&gt;e&lt;/sub&gt;</td>
<td>1.38 ± 0.88&lt;sup&gt;b&lt;/sup&gt;</td>
<td>N.A.</td>
<td>0.80 ± 0.01&lt;sup&gt;a&lt;/sup&gt;</td>
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<th>site B.</th>
<th>Parameters</th>
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<th>Groundwater</th>
<th>Ephemeral Wetland</th>
<th>Connected Wetland</th>
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</thead>
<tbody>
<tr>
<td>SUVA</td>
<td>1.81 ± 0.21&lt;sup&gt;a&lt;/sup&gt;</td>
<td>1.15 ± 0.43&lt;sup&gt;a&lt;/sup&gt;</td>
<td>1.87 ± 0.78&lt;sup&gt;a&lt;/sup&gt;</td>
<td>1.66 ± 0.53&lt;sup&gt;a&lt;/sup&gt;</td>
<td></td>
</tr>
<tr>
<td>E2/E3&lt;sub&gt;0.69&lt;/sub&gt;</td>
<td>4.00 ± 0.43&lt;sup&gt;b&lt;/sup&gt;</td>
<td>N.A.</td>
<td>3.69 ± 0.40&lt;sup&gt;a&lt;/sup&gt;</td>
<td>4.11 ± 0.76&lt;sup&gt;a&lt;/sup&gt;</td>
<td></td>
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<tr>
<td>S&lt;sub&gt;e&lt;/sub&gt;</td>
<td>0.75 ± 0.06&lt;sup&gt;b&lt;/sup&gt;</td>
<td>N.A.</td>
<td>0.71 ± 0.12&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.76 ± 0.10&lt;sup&gt;a&lt;/sup&gt;</td>
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</table>

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<tr>
<th>site C.</th>
<th>Parameters</th>
<th>Stream</th>
<th>Groundwater</th>
<th>Ephemeral Wetland</th>
</tr>
</thead>
<tbody>
<tr>
<td>SUVA&lt;sub&gt;0.69&lt;/sub&gt;</td>
<td>2.62 ± 0.26&lt;sup&gt;b&lt;/sup&gt;</td>
<td>1.17 ± 0.70&lt;sup&gt;a&lt;/sup&gt;</td>
<td>4.56 ± 1.23&lt;sup&gt;a&lt;/sup&gt;</td>
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<tr>
<td>E2/E3&lt;sub&gt;0.69&lt;/sub&gt;</td>
<td>3.39 ± 0.48&lt;sup&gt;b&lt;/sup&gt;</td>
<td>2.81 ± 0.90&lt;sup&gt;a&lt;/sup&gt;</td>
<td>4.03 ± 0.49&lt;sup&gt;a&lt;/sup&gt;</td>
<td></td>
</tr>
<tr>
<td>S&lt;sub&gt;e&lt;/sub&gt;</td>
<td>0.69 ± 0.10&lt;sup&gt;b&lt;/sup&gt;</td>
<td>1.06 ± 0.29&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.70 ± 0.10&lt;sup&gt;a&lt;/sup&gt;</td>
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Fig. 4. Fluorescence excitation-emission spectra of waters collected from stream (a), groundwater (b), and wetland (c) at site A in January 2012.
ture became obvious in warmer conditions (i.e., April). A higher temperature, with a range of 1–5 °C, was generally recorded in wetlands compared to stream and groundwater. Salt accumulation, as indicated with EC25, also became obvious at the end of season. Consider the smallest wetland in site A as an example. Its surface area was 6.75 m² and water depth was 0.4 m, and the water volume was about 2.7 m³. When the specific heat capacity of water is 4.1885 J g⁻¹ K⁻¹ and the solar energy dose of one full, clear, summer day at latitude 36° N is 14,031 kJ m⁻¹, the temperature could raise 8.4 °C in a day (without considering evaporation and other heat exchange processes). A higher water temperature can enhance water evaporation, drying, and salt accumulation. Therefore, the EC25 of an ephemeral wetland generally increased at the end of the wet season. For example, an EC25 of over 60 µS cm⁻¹ was recorded in the ephemeral wetland site B, three times higher than the level recorded in February (~20 µS cm⁻¹; Fig. S1-4b). In addition, air-water exchange in the static water condition of these ephemeral wetlands was minimal. With relatively rich nutrient inputs from foliar litters, low DO below 50% saturation was commonly observed.

The surrounding vegetation may also affect water quality because foliar litter is considered as an important nutrient source of isolated wetlands (Battle and Golladay, 2007; Crawford et al., 2007; Chow et al., 2013). In this study, the lowest DOC concentration (1.74 ± 0.11 mg L⁻¹) and SUVA (1.87 ± 0.78 L mg⁻¹ m⁻¹) were observed in the ephemeral wetland at site B, which was located in an open area without canopy coverage. Input of foliar litters to this wetland system was minimal. In contrast, the two wetlands in sites A and C were surrounded with deciduous trees, resulting in greater DOC concentrations released from foliar litters in the wetlands and thus greater C/N ratios.

4.2. Biogeochemical characteristics

The quality and quantity of foliar litters in a wetland are important factors influencing water quality and biogeochemical processes (Rejmankova and Houdkova, 2006; Bastviken et al., 2007). Lower pH and DO were observed in the ephemeral wetlands compared to their adjacent water bodies, and may be attributed to the decomposition of foliar litter (Deano and Robinson, 1985; Chow et al., 2013). Different carbon fractions released from decaying leaves increased the DOC concentration and C/N ratio in the wetland waters (Currie and Aber, 1997; Chow et al., 2011). Some of these degradable compounds such as polysaccharides and protein were found to enhance microbial activities, resulting in low DO in the wetlands (Howitt et al., 2007). Optical properties analyzed using SUVA and FRI showed that water contained significant portions of humic-acid-like and fulvic-acid-like compounds (Zhou et al., 2013). Dissolved organic nitrogen (DON) was the dominant N species (>50%) followed by NH₄-N, suggesting N mineralization or nitrification was minimal within the wetlands.

Most of the litter inputs occurred prior to January but the breakdown of these organic matters through invertebrate or microbial decomposition could have been limited due to the low temperature condition in winter time. From January to March, the nutrient concentration, including DOC and TDN were relatively stable and no obvious changes occurred in the three months. However, a decrease of humic-acid-like and fulvic-acid-like fractions was observed from January to March, but an increase of aromatic protein and microbial byproduct-like fractions in the ephemeral wetlands was observed (Fig. 5). The changes in DOM composition but not quantity could probably be attributed to the effects of temperature and sunlight (Chow et al., 2006; Chow et al., 2013). In January, temperature was generally <10 °C and the biological decomposition should be minimal. In contrast, opened canopy after litter fall allowed sunlight penetration onto the wetlands. DOM could be degraded from large aromatic molecules to small water soluble non-aromatic compounds through photobleaching processes (Brinkmann et al., 2003; Chow et al., 2013). Lower SUVA but higher E2/E3 values in waters collected in January were generally observed when comparing to those collected in March. With a shallow water depth, photochemical processes could be an important process controlling nutrient cycling within the ephemeral wetlands in winter time (Piccini et al., 2009).

4.3. Soil infiltration

Changes in water levels (ΔH/Δt) have been used to characterize hydrological connectivity of riparian wetlands with adjacent water bodies (Cabezas et al., 2011). The rapid and simultaneous changes in water levels during rainstorm events suggested the hydrological connectivity of the ephemeral wetlands with the surrounding water bodies. Considering site A as an example (Fig. 2), sudden increase of water levels in groundwater and wetland were observed with intensive rainstorms (>0.5 mm per day). Notably, the water levels promptly dropped back to their background levels in only 1–2 days. Water loss of a completely isolated wetland should be through evaporation only and their water levels should be relatively stable in such low temperature condition (T < 15 °C during the study period). The decrease in water levels in such a short time indicated that water seeped out to the surrounding areas rather than being lost due to evaporation. In fact, the water level peaks of wetlands closely matched the peaks of groundwater. The strong correlations in ΔH/Δt among water bodies further support this argument (Tables SI-1–SI-3).

In spite of their potential hydrological connectivity, the water quality of the ephemeral wetlands differed from their adjacent water bodies. Distinguishable temperature, pH, DO, DOM, and nutrients were found among different water bodies (Tables 1 and 2 and SI-4). Such biogeochemical disconnection among water bodies could occur over even a short distance as long as there was no direct surface water connection. For example, the ephemeral wetland in site A was 10.7 m away from the 1st order stream. Despite the short distance between the ephemeral wetland and the 1st order stream and the hydrological connectivity suggested by the changes in water level (i.e. ΔH/Δt), their water chemistry was significantly different from each other. The biogeochemical disconnection was attributed to the soil infiltration process. Soil between the two water bodies served as a chemical filter that could absorb and reduce nutrients and minimize the chemical mixing between two water bodies (Parsons et al., 2004; Chow et al., 2009; Schmidt et al., 2012). Furthermore, soil has been demonstrated to
effectively remove organic matter from water through filtration processes (Maeng et al., 2008; Mudarra et al., 2011). Soil acting as sorbents can absorb certain DOM fractions, particularly hydrophobic fractions, and reduce the DOC concentration passing through the soil columns. A significant decrease in UV/VIS absorbance from wetland water to groundwater was recorded. FRI and SUVA analyses also demonstrated that the humic acid-like compounds, the major fraction found in wetlands, were not observed in groundwater (Fig. 5).

In addition to serving as a chemical filter, soil can serve as a bioreactor altering the biogeochemical processes and nutrient cycling such as natural organic matter degradation and denitrification (Diem et al., 2013). Consider N species along a potential water connection (i.e., wetland ←→ groundwater ←→ stream). Wetlands were dominated with dissolved organic nitrogen (DON) because of leachate from foliar litter. Microbial processes in the soil could convert DON into inorganic N through mineralization when wetland waters seeped to groundwater (Wolf et al., 2013). As shown in groundwater in site C, it was dominated by inorganic N with similar portions of NH₄-N and NO₃-N. This inorganic N was further processed through nitrification when water exported to the stream (Racchetti et al., 2011). As seen in the 2nd order stream in site C, nitrate was the only dominant species. Differences in N species demonstrated that ephemeral wetlands, groundwater, and the stream had unique water chemistry or were biogeochemically distinct in spite of their proximity.

5. Conclusion

Water chemistry and physical parameters of three ephemeral wetlands and their adjacent water bodies in the Piedmont ecoregion were monitored during their flooded period in 2012. Nutrient and water quality analyses indicated their chemistries differed. Lower pH and DO were observed in ephemeral wetlands we believe because of foliar litter decomposition, while higher EC₂₅ and temperature fluctuations were recorded most likely because smaller wetlands were less buffered against environmental changes. A higher DOC concentration with a significant portion of humin-acid-like and fulvic-acid-like compounds was found in ephemeral wetlands. Also, dissolved organic N was the dominant N species in wetlands. Soil infiltration processes most likely altered the water chemistry, reducing the DOC concentration and converting organic nitrogen into inorganic nitrogen in groundwater and stream water. The groundwater connections coupled with the lack of biogeochemical similarity among the ephemeral wetlands and the permanent water bodies suggest that the ephemeral wetlands not only contribute to groundwater recharge, but soils surrounding the ephemeral wetlands contribute to the maintenance of the chemical integrity of the groundwater.

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Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.ecoleng.2016.06.075.

References


Helms, J.R., Stubbins, A., Ritchie, J.D., Minor, E.C., Kieber, D.J., Mopper, K., 2008. Absorption spectral slopes and slope ratios as indicators of molecular weight,


